

## PHENOTYPIC VARIATION IN RETAIL BEEF YIELD IN ANGUS CATTLE

K.A. Donoghue<sup>1</sup>, L.M. Cafe<sup>2</sup>, B.J. Walmsley<sup>3</sup> and C. Duff<sup>4</sup>

<sup>1</sup>NSW Department of Primary Industries, Agricultural Research Centre, Trangie, NSW, 2823

<sup>2</sup>NSW Department of Primary Industries, Livestock Industries Centre, Armidale, NSW, 2351

<sup>3</sup>Animal Genetics & Breeding Unit\*, University of New England, Armidale, NSW, 2351 Australia

<sup>4</sup>Angus Australia, 86 Glen Innes Road, Armidale, NSW, 2350

### SUMMARY

This paper reports preliminary estimates of phenotypic variation and relationships for carcass traits using records from 386 Angus steers. Carcass traits recorded included carcass weight (CWT), retail beef yield (RBY), carcass intramuscular fat (CIMF), carcass P8 fat (CP8), carcass rib fat (CRIB) and carcass eye muscle area (CEMA). Carcass trait averages for animals used in this study were higher than for those used for the previous estimation of RBY parameters, and the differences evident between summary statistics for historic and contemporary carcass traits records highlights the value of continually recording carcass traits, as well as the value of re-examining relationships between carcass traits. Phenotypic correlations between carcass traits indicated that animals with higher RBY also had, phenotypically, heavier CWT ( $r_p = 0.12$ ), larger CEMA ( $r_p = 0.31$ ), lower CRIB and CP8 ( $r_p = -0.28$  and  $-0.10$  respectively) and lower CIMF ( $r_p = -0.16$ ). For every 1-kg increase in RBY EBV, recorded CWT, RBY and CEMA were greater by 2.08kg, 0.24% and 2.77cm<sup>2</sup>, respectively. In contrast, for every 1-kg increase in RBY EBV, recorded CIMF, CRIB and CP8 were lower by 0.39%, 1.31mm and 1.81mm, respectively. These preliminary results highlight the importance of continually recording carcass traits as well as the value of periodically re-examining relationships between carcass traits.

### INTRODUCTION

Retail Beef Yield (RBY) is a key driver of profitability in the Australian beef industry, and the RBY EBV underpins BreedObject \$Indices which allow selection for profitability in the major beef breeds. However, in current genetic evaluations, the trait of RBY is estimated almost exclusively from correlations with live weight, live animal scanned eye muscle area and fat depths. This is due to very limited recording of abattoir carcass data and specifically, RBY phenotypes. RBY phenotypes are not generated routinely because accurate data is time-consuming and expensive to collect. Previous estimates of phenotypic and genetic parameters for RBY were undertaken on a relatively small number of animals (n=1,930) born between 1994 and 2000 (Reverter *et al.* 2000; Reverter *et al.* 2003). Genetic improvement in carcass traits in the 20 years since the last RBY records were collected has led to significant improvements in carcass traits (Johnston 2007).

The purpose of this project is to generate RBY phenotypes on 1,000 fully pedigreed, genotyped and genetically described Angus steers. These will be used to re-estimate BREEDPLAN RBY parameters for incorporation into BREEDPLAN analyses to provide RBY EBVs. Animals in the project will be measured for a comprehensive suite of traits from birth until slaughter, including live weight and scan data taken from weaning until feedlot exit, and these traits will be assessed for their relationship with RBY. This will provide information to determine the effectiveness of exploiting early in life measurements as genetic indicators for RBY which may reduce the need to record actual RBY. The collection of RBY data will provide a reference population on which genomic information will be applied to provide RBY EBVs. Data used in this study comprises animals from the first two cohorts (of four) of the RBY project and provides preliminary estimates of phenotypic relationships between RBY and other carcass traits.

\* A joint venture of NSW Department of Primary Industries and the University of New England

## **MATERIALS AND METHODS**

The animals used in this project were steers from the NSW DPI Trangie and Glen Innes research herds born in 2015 (n=150) and 2016 (n=236). All steers were fully pedigreed and, in collaboration with the Angus Sire Benchmarking Program had a comprehensive suite of traits measured from birth, based on BREEDPLAN collection protocols, and genomic testing (GGP-LD or Angus GS profile) conducted. There were 109 sires represented in the dataset, with the number of progeny for each sire ranging from 1 to 7. Fifty three of the 109 sires had progeny represented in both herds (Trangie and Glen Innes), and these 53 sires accounted for 58% (224/386) of the data.

Following weaning on the birth property, all steers were backgrounded on irrigated ryegrass /oats pasture and other pastures at Grafton Primary Industries Institute and were grain finished at UNE Tullimba Research Feedlot for 150 days to fit the John Dee, Warwick Angus Gold Grid. The steers were slaughtered in lots of approximately 50 animals at John Dee, Warwick, to suit the number of sides which can be boned for out for RBY at the facility in one shift. The cohorts were split into kill groups to ensure the integrity of the contemporary groups and based on balance for sire and live weight.

Following commercial AUS-MEAT carcass preparation (Anon. 2007), carcasses were weighed and hot P8 fat depth recorded. Fat trim following hot standard carcass weight measurement was restricted over the 12/13 rib to allow meaningful MSA rib fat data collection, and standard excess fat trim was conducted on the remainder of the carcass. Carcasses were tagged and chilled overnight, and Meat Standards Australia (MSA) carcass grading data (Anon. 2008) was collected by registered MSA graders on the right hand sides the following morning prior to boneout. A sample of the cube roll was collected for lab analysis to determine IMF% by the Meat Science Department at the University of New England following the near infrared spectrophotometry (NIR) method described by Perry *et al.* (2001).

For boneout a standard set of AUS-MEAT boneless primals, standard trim of 10mm fat, and estimated chemical lean of trim of 65% was conducted. The primal cuts included in the measure of RBY were: cube roll, chuck, chuck tender, blade, brisket, intercostals, FQ shin/shank, rib end meat, inside skirt, FQ trim, topside/inside, thick flank, rump, striploin, tenderloin, thin flank, silverside, HQ shin/shank, HQ trim. Carcass components of FQ fat trim, FQ bone, HQ fat trim and HQ bone did not contribute to RBY. Boning room pre-trim was weighed for completeness, though this was usually only 50 to 200g of fat and added to the weight of fat trim. The cold weight of the sides was measured prior to quartering and entering the boning room. This cold weight was used as a measure of reliability of the RBY data, as the recovery of this weight should be close to 100% and should be consistent across sides.

Data available for the analyses included carcass weight (CWT), eye muscle area (CEMA), retail beef yield percentage (RBY), intramuscular fat percentage (CIMF), subcutaneous fat depths at the P8 rump (CP8) and the 12<sup>th</sup> and 13<sup>th</sup> rib (CRIB) sites.

**Model of analysis.** Contemporary group for carcass records included effects of herd of origin and kill date. Variance and covariance components were estimated with an animal model using ASReml (Gilmour *et al.* 2015). A fixed effect of contemporary group, random direct genetic effects, and residual effects were included in the standard model. Univariate analyses were undertaken to obtain phenotypic variances for each trait, while pairwise bivariate analyses were undertaken to obtain phenotypic correlations. Regression of recorded carcass traits on the corresponding BREEDPLAN carcass EBVs were undertaken using PROC REG (SAS). Carcass traits were adjusted as per standard BREEDPLAN analyses (CWT adjusted to 750 days and all other traits adjusted to 400 kg carcass weight) for the regression analyses. BREEDPLAN carcass EBVs on animals were obtained prior to the inclusion of carcass data from the project in the genetic evaluation.

**RESULTS AND DISCUSSION**

Summary statistics for the carcass traits are shown in Table 1. Most carcass trait averages for animals used in the previous estimation of RBY parameters were markedly lower than those presented in Table 1 (From Reverter *et al.* 2003: Slaughter age = 630±128 days; CWT= 269±55 kg; RBY=67.03±3.69%; CIMF=4.64±2.23%; CP8=10.19±4.57 mm; CRIB=8.21±4.45 mm; CEMA=81.59±15.9 cm<sup>2</sup>). The differences evident between summary statistics for historic and contemporary carcass trait records highlight the value of recording carcass traits in modern cattle populations, as well as the value of re-examining relationships between carcass traits in this population.

**Table 1. Descriptive statistics for carcass traits**

Trait	No. records	Average (SD)	Minimum	Maximum	$\sigma_p^2$ (SE)
Age at slaughter (days)	386	596.4 (21.8)	536.0	635.0	-
CWT (kg)	386	344.7 (30.1)	269.0	428.5	889.10 (72.96)
RBV (%)	386	73.84 (2.02)	68.69	78.92	1.59 (0.12)
CIMF (%)	386	6.29 (2.04)	2.01	13.99	2.91 (0.22)
CP8 (mm)	386	16.24 (4.18)	10.00	30.00	13.73 (1.04)
CRIB (mm)	386	12.58 (4.45)	4.00	28.00	11.09 (0.81)
CEMA (cm <sup>2</sup> )	386	81.48 (8.44)	57.00	108.0	37.27 (2.89)

Phenotypic correlations ( $r_p$ ) between carcass traits were similar to those reported in the literature for the same traits in temperate breeds (Reverter *et al.* 2003) for the majority of traits (Table 2). Of those that were different to previous estimates, the majority of  $r_p$  were in the same direction but of a different magnitude;  $r_p$  between RBV and CIMF (-0.16 vs -0.31 in literature);  $r_p$  between RBV and CP8 (-0.10 vs -0.40 in literature);  $r_p$  between RBV and CEMA (0.31 vs 0.14 in literature). The remaining estimates that indicated relationships in a different direction to previous estimates are most likely a reflection of the relatively low numbers analysed so far in this project;  $r_p$  between CWT and CIMF (0.12 vs -0.19 in literature);  $r_p$  between CWT and CP8 (0.15 vs -0.19 in literature);  $r_p$  between CWT and CRIB (0.19 vs -0.19 in literature).

**Table 2. Phenotypic correlations (SE) between carcass traits**

Trait	RBV	CIMF	CP8	CRIB	CEMA
CWT	0.12 (0.05)	0.12 (0.06)	0.15 (0.05)	0.19 (0.05)	0.38 (0.05)
RBV		-0.16 (0.05)	-0.10 (0.05)	-0.28 (0.05)	0.31 (0.05)
CIMF			0.07 (0.05)	0.06 (0.05)	-0.14 (0.05)
CP8				0.25 (0.05)	-0.09 (0.05)
CRIB					-0.17 (0.05)

Phenotypic correlations between carcass traits would indicate that animals with higher RBV also had, phenotypically, heavier CWT ( $r_p = 0.12$ ), larger CEMA ( $r_p = 0.31$ ), lower CRIB and CP8 ( $r_p = -0.28$  and  $-0.10$  respectively) and lower CIMF ( $r_p = -0.16$ ). In summary, results so far would indicate that phenotypic relationships between recorded carcass traits follow the pattern expected, given previous estimates, and make sense biologically. This confirms the quality of the data that is being collected and highlights the value of the data to estimate updated genetic parameters for these traits once more records have been obtained.

Regression coefficients from regression of recorded carcass traits on corresponding BREEDPLAN carcass EBVs are presented in Table 3. These results indicate that, on average, for every 1-kg increase in RBY EBV, recorded CWT, RBY and CEMA were greater by 2.08kg, 0.24% and 2.77cm<sup>2</sup>, respectively. In contrast, for every 1-kg increase in RBY EBV, recorded CIMF, CRIB and CP8 were lower by 0.39%, 1.31mm and 1.81mm, respectively.

**Table 3. Regression coefficients from regression of recorded carcass traits on corresponding BREEDPLAN carcass EBVs**

Trait	CWT EBV	RBY EBV	CIMF EBV	CP8 EBV	CRIB EBV	CEMA EBV
CWT	2.11**	2.08	7.22	-5.26**	-5.44**	-0.25
RBY	-0.01*	0.24**	-0.08	-0.05	-0.07	0.13**
CIMF	0.006	-0.39**	1.31**	0.08	0.10	-0.07
CP8	-0.05**	-1.81**	-0.22	1.17**	1.27**	-0.26**
CRIB	-0.03	-1.31**	-0.10	0.70**	1.00**	-0.33**
CEMA	-0.04	2.77**	-1.67**	-0.52**	-0.71**	1.31**

\* Significance at P<0.05; \*\* Significance at P<0.001

## CONCLUSIONS

In the majority of cases, phenotypic correlations and regression coefficients were such that relationships between carcass traits were in the expected direction and similar in magnitude to literature estimates. As the number of carcass records increase as this project continues it is anticipated that genetic parameters for carcass traits will be estimated. This will allow more detailed investigations into the appropriateness of the current BREEPLAN carcass genetic parameters for contemporary Angus cattle.

## ACKNOWLEDGEMENTS

This work was funded by NSW DPI and Angus Australia through the MLA Donor Company program. The capable assistance of staff at Glen Innes Agricultural Research and Advisory Station, Trangie Agricultural Research Centre, Grafton Primary Industries Institute and Angus Australia is gratefully acknowledged. The management and staff at John Dee, Warwick abattoir provided much capable assistance during collection of the carcass data.

## REFERENCES

- Anon. (2007) 'AUS-MEAT National Accreditation Standards' 2007 Edition (AUS-MEAT Ltd, Brisbane).  
 Anon. (2008) *Aust. J. Exp. Agr.* **48**: 1360.  
 Gilmour A.R., Gogel B.J., Cullis B.R., Welham S.J. and Thompson R. (2015) 'ASReml User Guide Release 4.1'. VSN International Ltd, Hemel Hempstead, UK.  
 Perry D., Shorthose W.R., Ferguson D.M. and Thompson J.M. (2001) *Aust. J. Exp. Agr.* **41**: 953.  
 Johnston D.J. (2007) *Proc. Assoc. Advmt. Anim. Breed. Genet.* **17**: 8.  
 Reverter A., Johnston D.J., Perry D., Goddard M.E. and Burrow H.M. (2003) *Aust. J. Agric. Res.* **54**: 119.  
 Reverter A., Johnston D.J., Graser H.-U., Wolcott M.L. and Upton W.H. (2000) *J. Anim. Sci.* **78**: 1786.